



Organización Mundial
de Sanidad Animal
Fundada como OIE

Vigilancia de la Resistencia Antimicrobiana: Bases y Herramientas Microbiológicas y Genómicas



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Tópicos

- Conceptos en RAM y producción.
- Bases microbiológicas/genéticas de la RAM.
- Antibióticos en acuicultura.
- Principales mecanismos de RAM.
- Bases genéticas de la RAM en acuicultura.
- RAM y Salud Única
- Conclusiones.

Conceptos en RAM y producción

- ✓ La microbiota es esencial para los seres vivos y ecosistemas.
- ✓ Esta microbiota es específica para cada especie animal.
- ✓ ATMs seleccionan y reducen la diversidad de la microbiota.
- ✓ La disminución de la microbiota favorece infecciones.
- ✓ El uso de ATM favorece la emergencia/transferencia horizontal de la RAM.

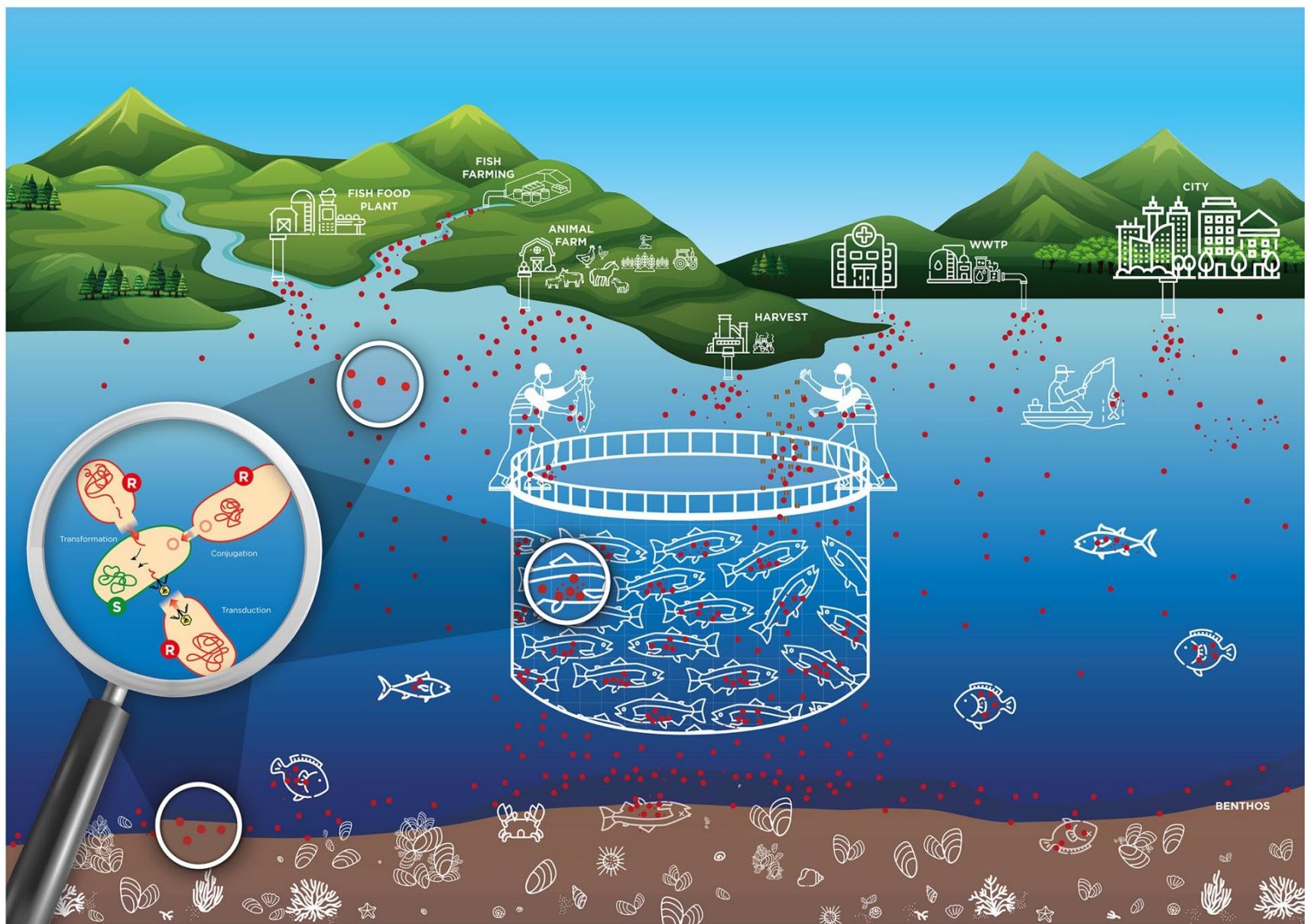
Conceptos en RAM y producción

- ✓ ATM en producción generan residuos.
- ✓ Alimentos derivados pueden contener residuos y/o bactérias resistentes.
- ✓ Bacterias en alimentos pueden transmitir AMR para patogenos o comensales humanos.
- ✓ Residuos son descartados para el ambiente, o reutilizados (abono orgánico).
- ✓ Ambientes acuáticos son *hotspots* para transferencia de genes RAM.

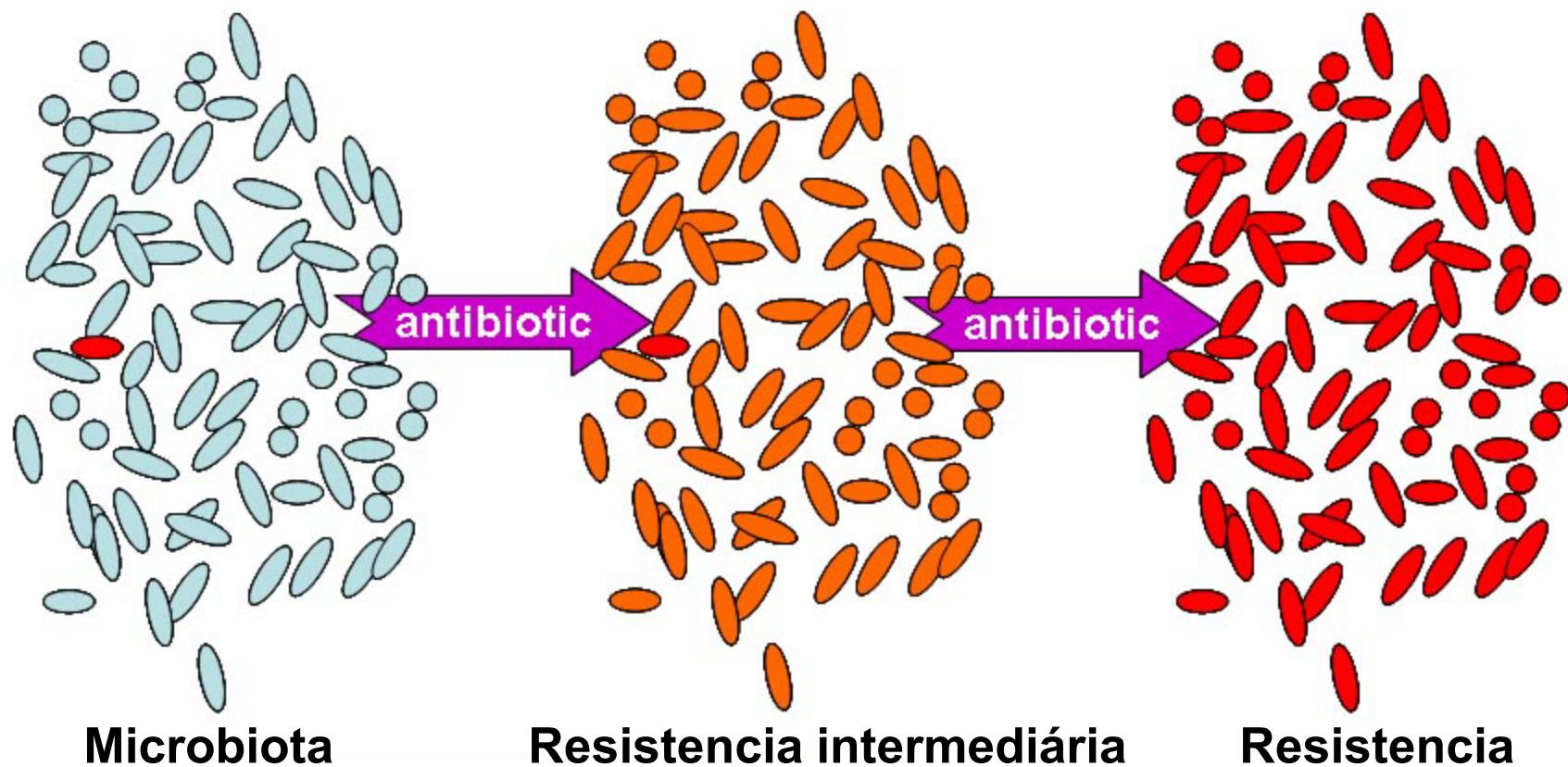
Dissemination of AMR



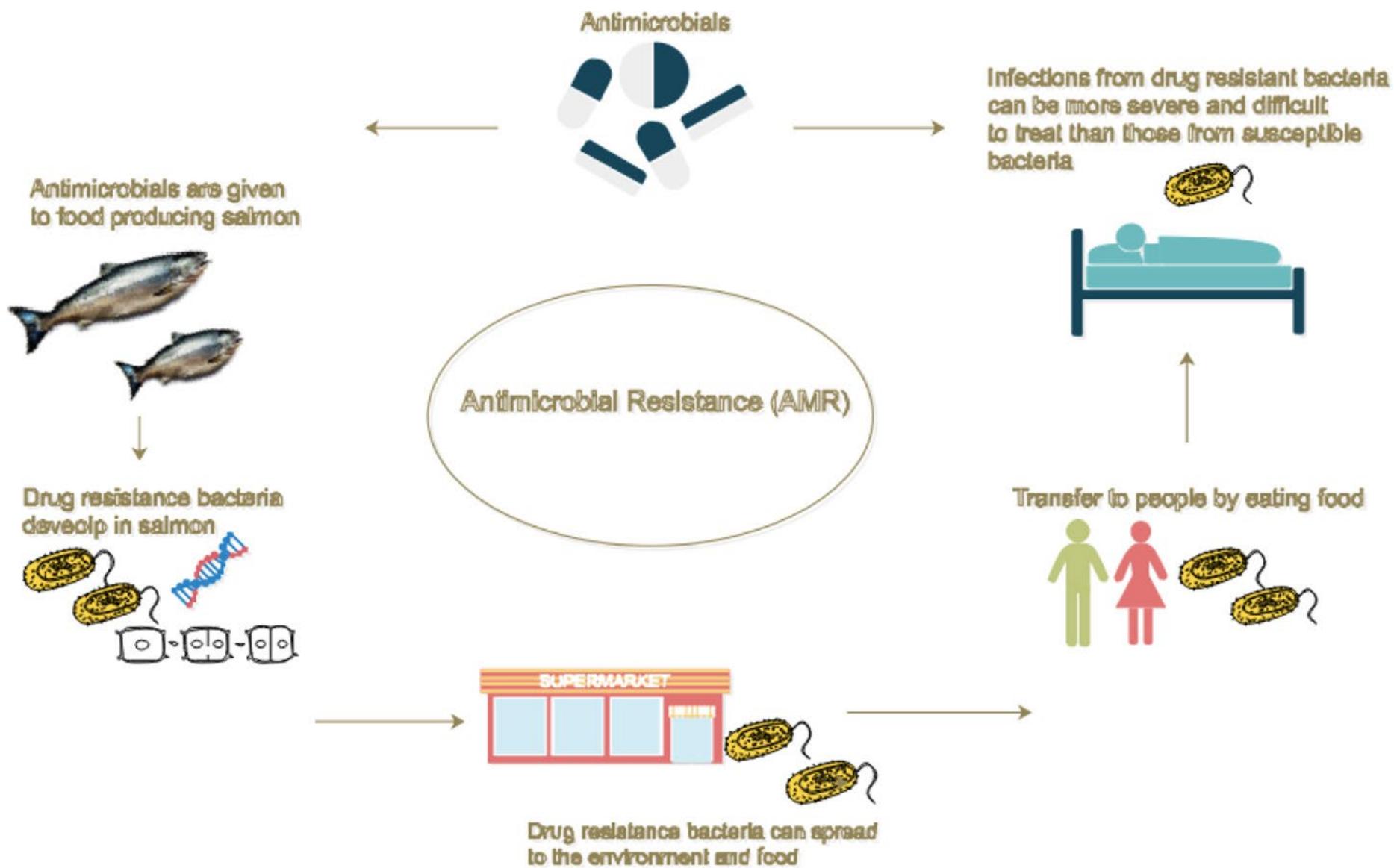
Origen de la RAM en ambientes acuáticos



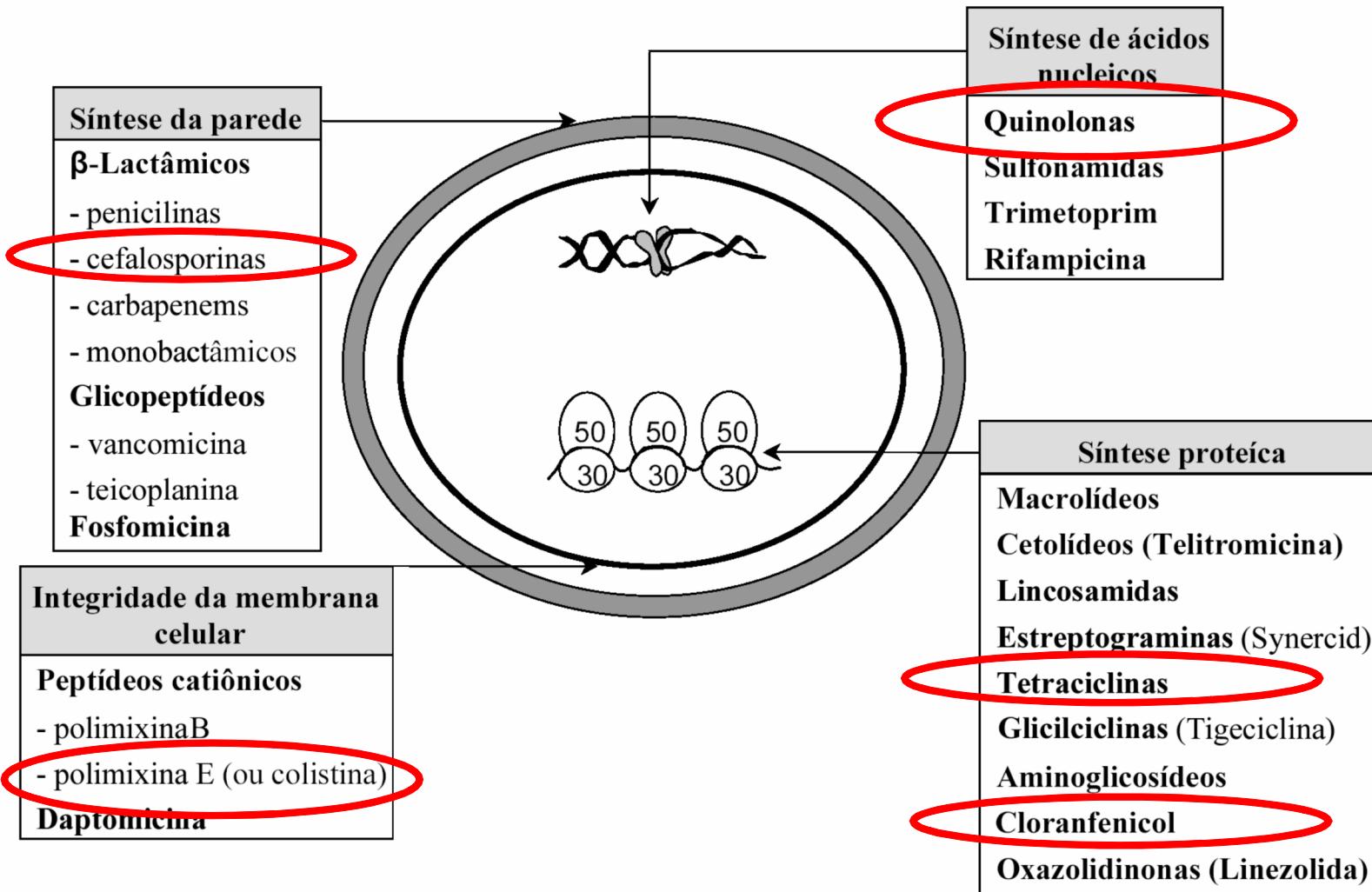
RAM y presión selectiva



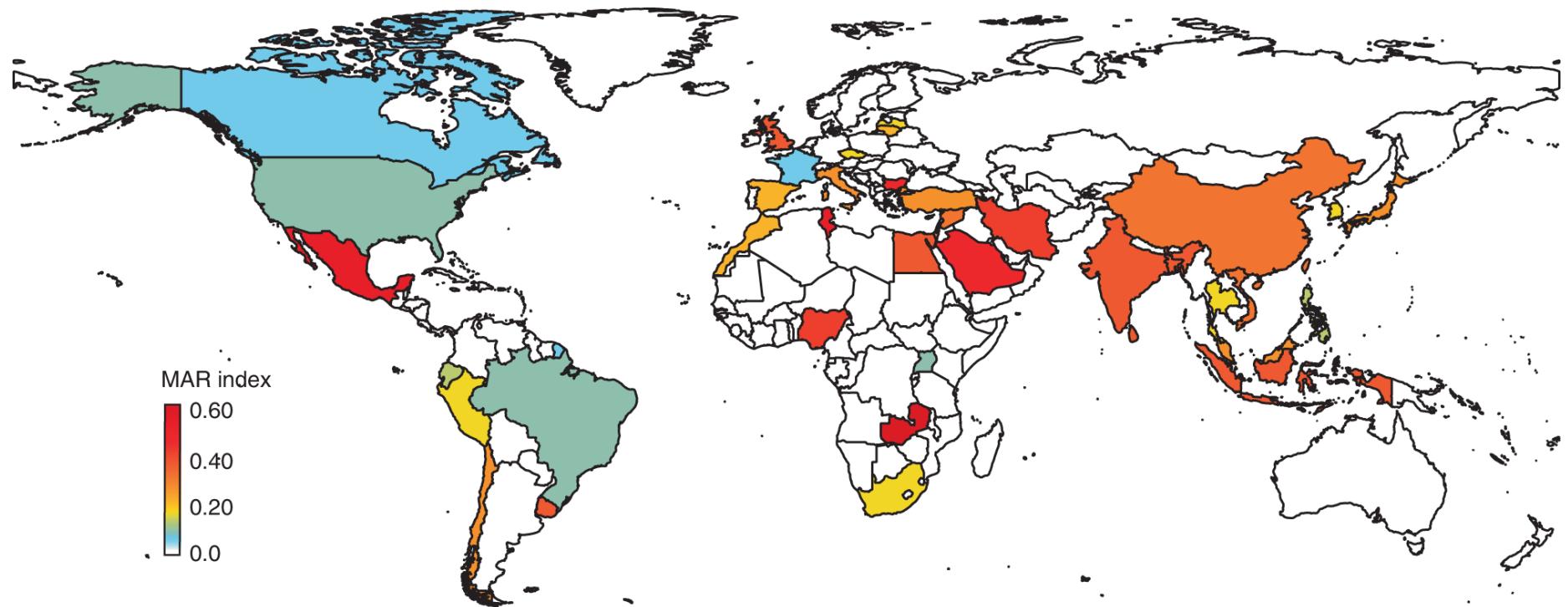
Importancia de la RAM en acuicultura



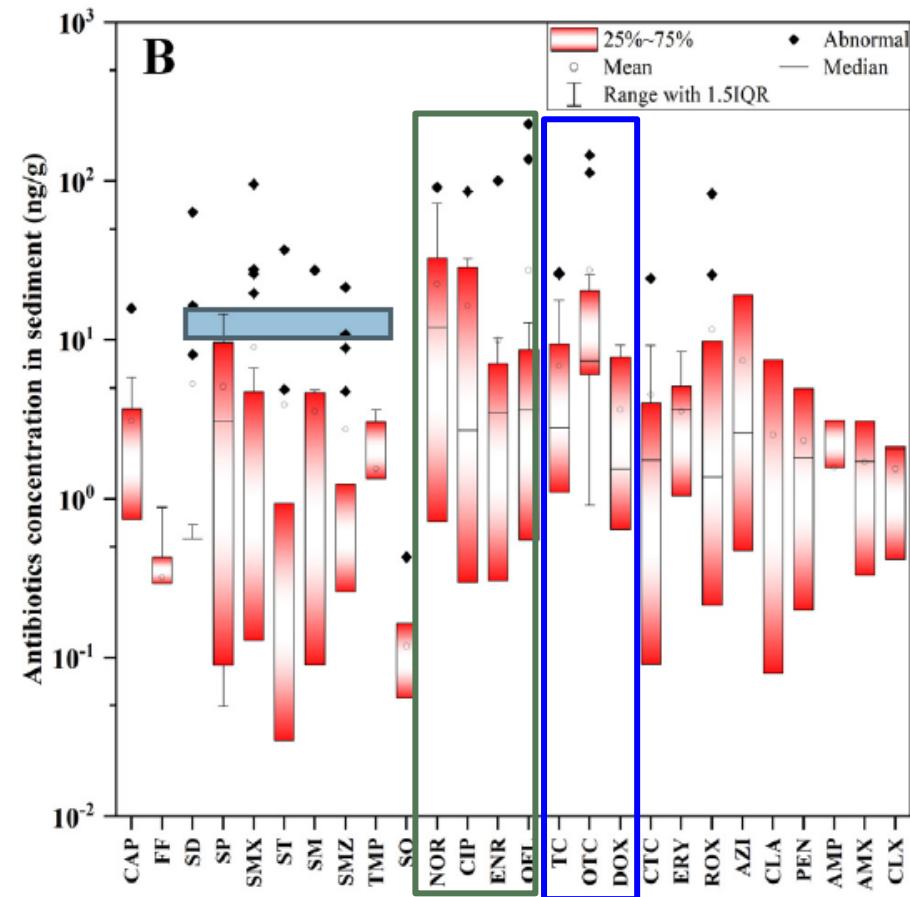
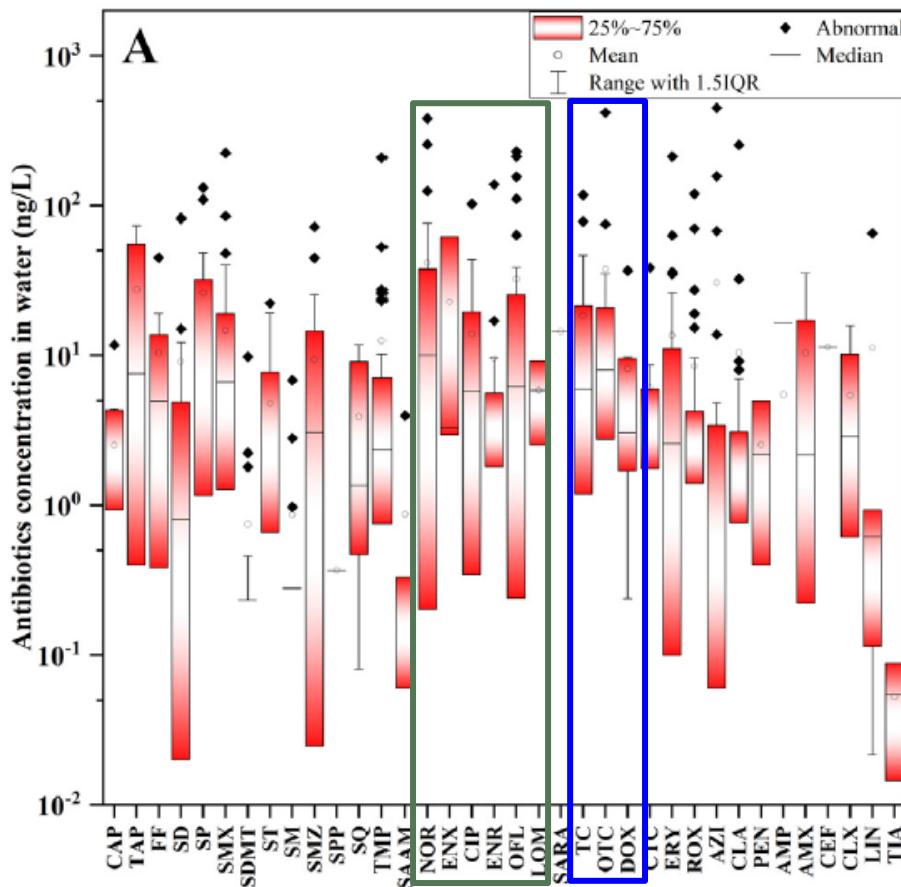
Antibacterianos en animales de producción



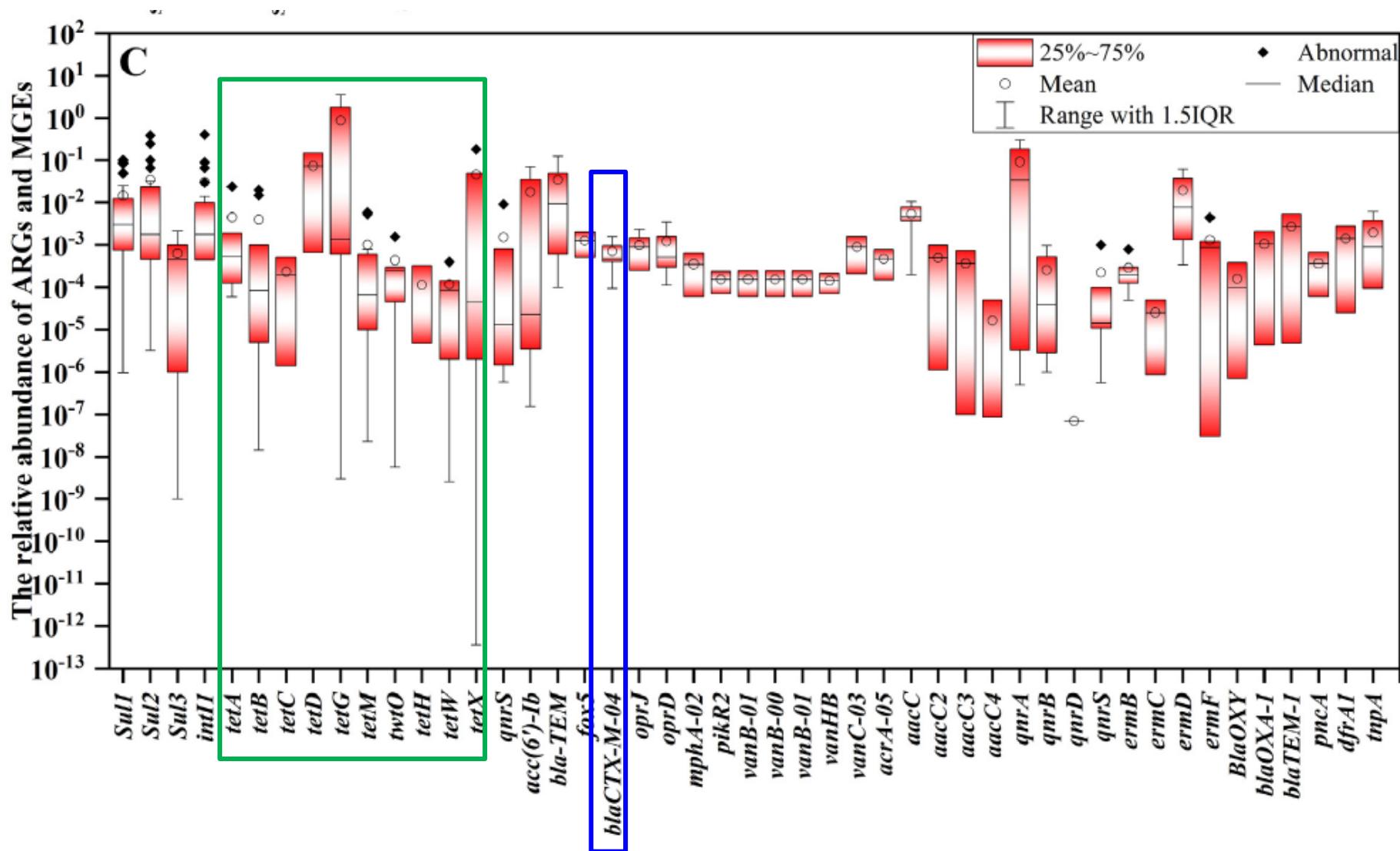
Índice RAM Global en acuicultura



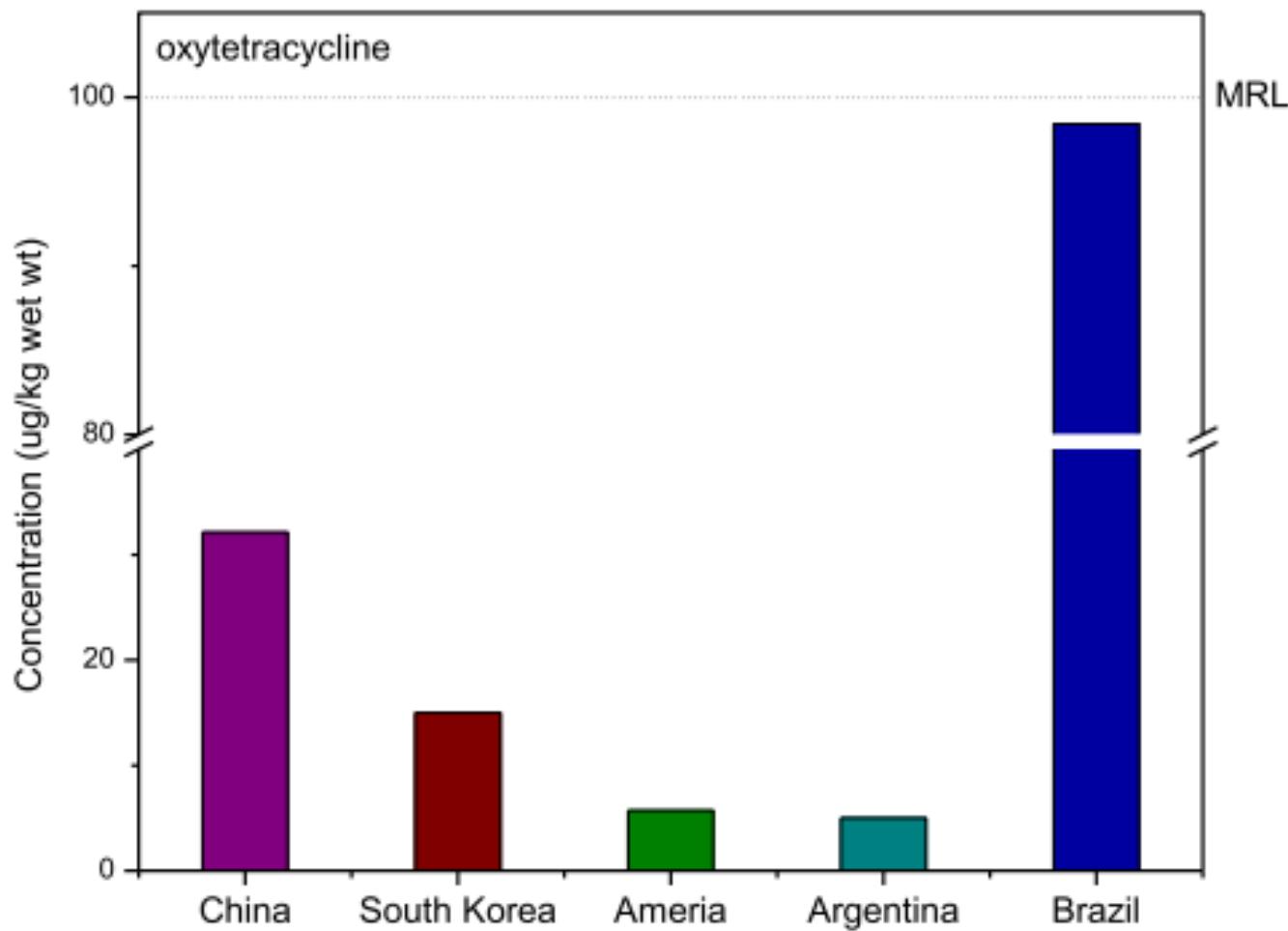
Concetración de ATB en ambiente estuarino/costero



Genes de resistencia en ambiente estuarino/costero

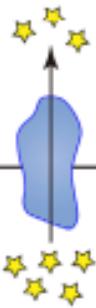


Oxitetraciclina en productos de acuicultura



Resistência a las tetraciclinas

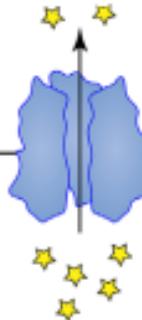
A1



Tetracycline-specific efflux pump genes (n=34)

<i>tet(A)</i>	<i>tet(B)</i>	<i>tet(C)</i>	<i>tet(D)</i>	<i>tet(E)</i>	<i>tet(G)</i>	<i>tet(H)</i>
<i>tet(J)</i>	<i>tet(K)</i>	<i>tet(L)</i>	<i>tet(V)</i>	<i>tet(Y)</i>	<i>tet(Z)</i>	<i>tet(30)</i>
<i>tet(31)</i>	<i>tet(33)</i>	<i>tet(35)</i>	<i>tet(38)</i>	<i>tet(39)</i>	<i>tet(40)</i>	<i>tet(41)</i>
<i>tet(42)</i>	<i>tet(43)</i>	<i>tet(45)</i>	<i>tet(57)</i>	<i>tet(58)</i>	<i>tet(59)</i>	<i>tet(62)</i>
<i>tetA(P)</i>	<i>tetAB(46)</i>	<i>tetAB(60)</i>	<i>otr(B)</i>	<i>otr(C)</i>	<i>tcr3</i>	

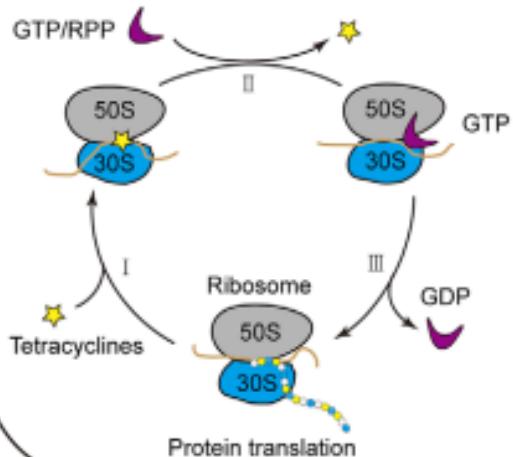
A2



Multidrug-resistant efflux pump genes (n=9)

<i>acrAB-toIC</i>	<i>acrEF-toIC</i>	<i>adeABC</i>	<i>efpA</i>	<i>mexAB-oprM</i>
<i>mexCD-oprJ</i>	<i>mexEF-oprN</i>	<i>mexXY-oprM</i>	<i>oqxAB-toIC</i>	

C



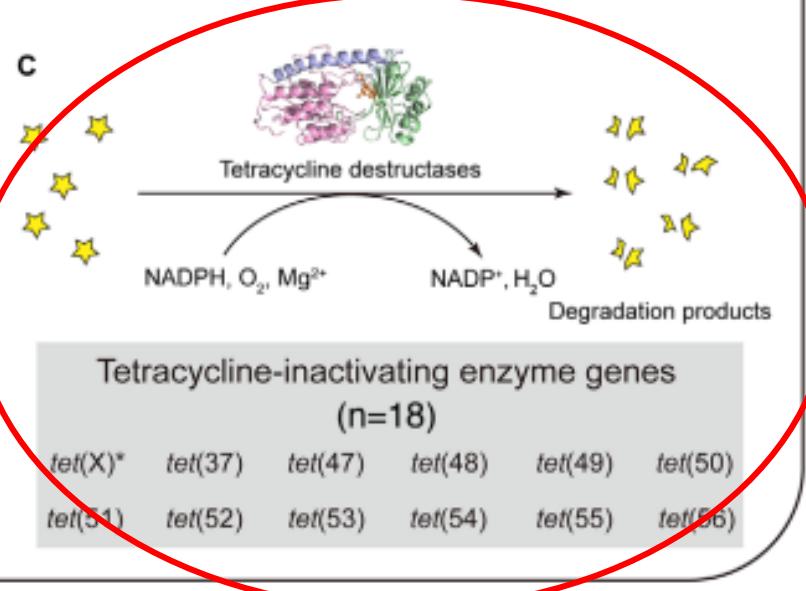
B

Ribosomal protection genes (n=13)

<i>tet(M)</i>	<i>tet(O)</i>	<i>tet(S)</i>
<i>tet(Q)</i>	<i>tet(T)</i>	<i>tet(W)</i>
<i>tet(32)</i>	<i>tet(36)</i>	<i>tet(44)</i>
<i>tet(61)</i>	<i>otr(A)</i>	<i>tetB(P)</i>
<i>tet</i>		

Tetracycline-inactivating enzyme genes (n=18)

<i>tet(X)*</i>	<i>tet(37)</i>	<i>tet(47)</i>	<i>tet(48)</i>	<i>tet(49)</i>	<i>tet(50)</i>
<i>tet(51)</i>	<i>tet(52)</i>	<i>tet(53)</i>	<i>tet(54)</i>	<i>tet(55)</i>	<i>tet(56)</i>





Article

Genetic Characterization of the Tetracycline-Resistance Gene *tet(X)* Carried by Two *Epilithonimonas* Strains Isolated from Farmed Diseased Rainbow Trout, *Oncorhynchus mykiss* in Chile

Christopher Concha ¹, Claudio D. Miranda ^{1,2,*}, Javier Santander ³ and Marilyn C. Roberts ⁴

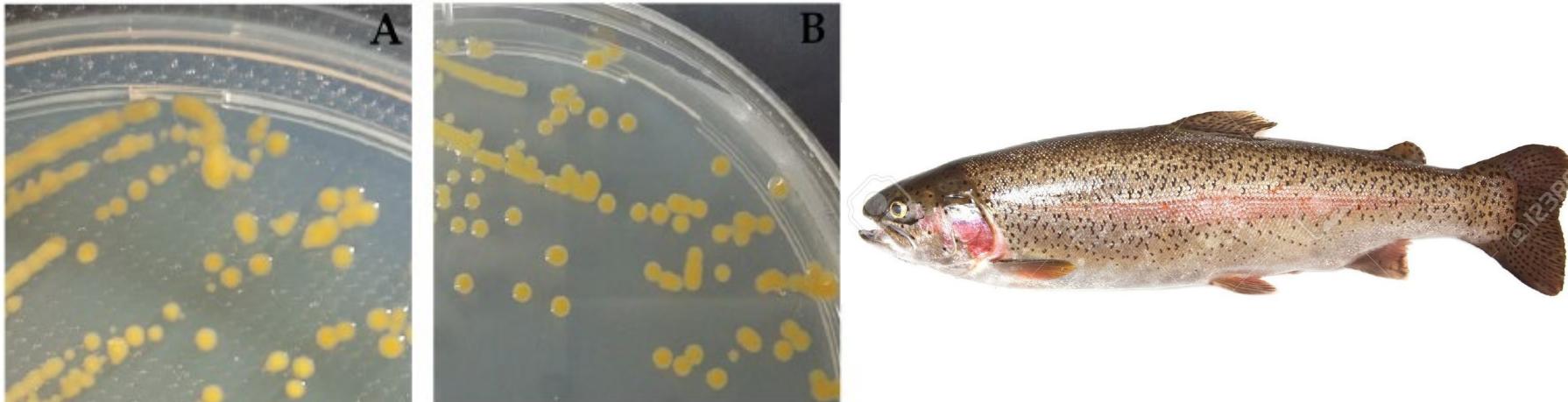
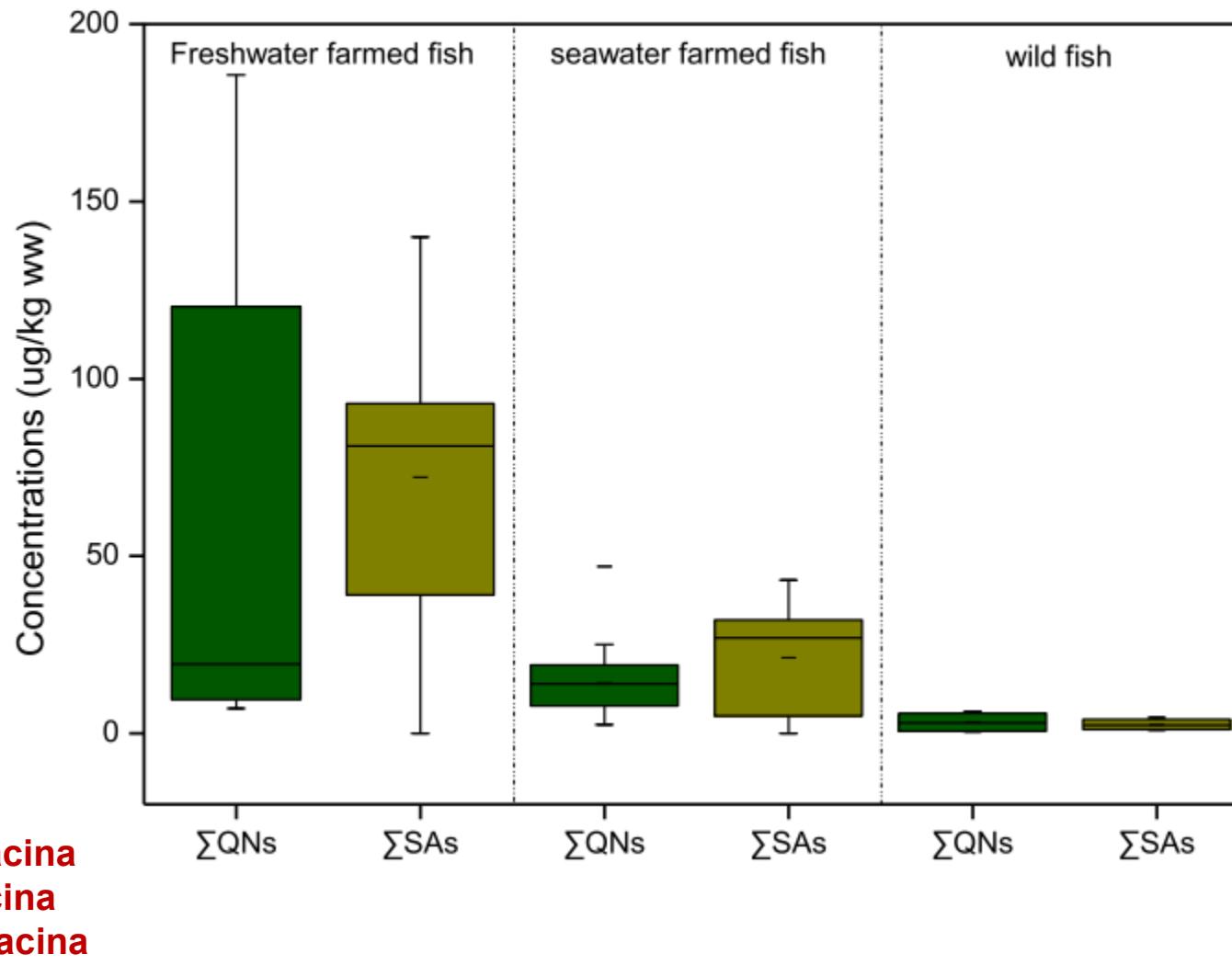


Figure 1. Colony morphotypes of the *Epilithonimonas* strains recovered from diseased rainbow trout from Chilean farms grown on TYES agar: (A) FP105; (B) FP211-J200.

Residuos de fluoroquinolonas y sulfas en pescados



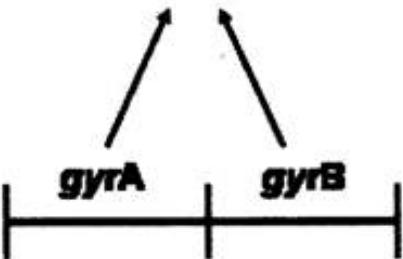
Resistencia a las fluoroquinolonas

A

Enrofloxacina
Ciprofloxacina



gyrase



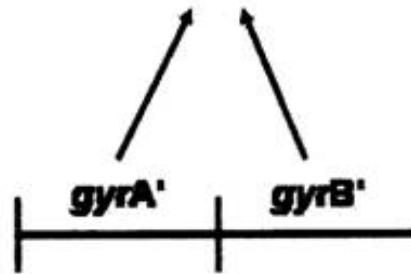
Bactéria Suscetível

B

Enrofloxacina
Ciprofloxacina



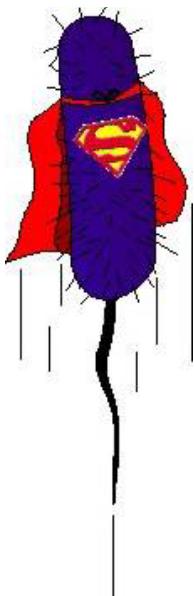
gyrase



Bactéria Resistente

Mutación

Patógenos de prioridad crítica



Priority 1: CRITICAL

- *Acinetobacter baumannii*, carbapenem-resistant
- *Pseudomonas aeruginosa*, carbapenem-resistant
- *Enterobacteriaceae*, carbapenem-resistant, ESBL-producing



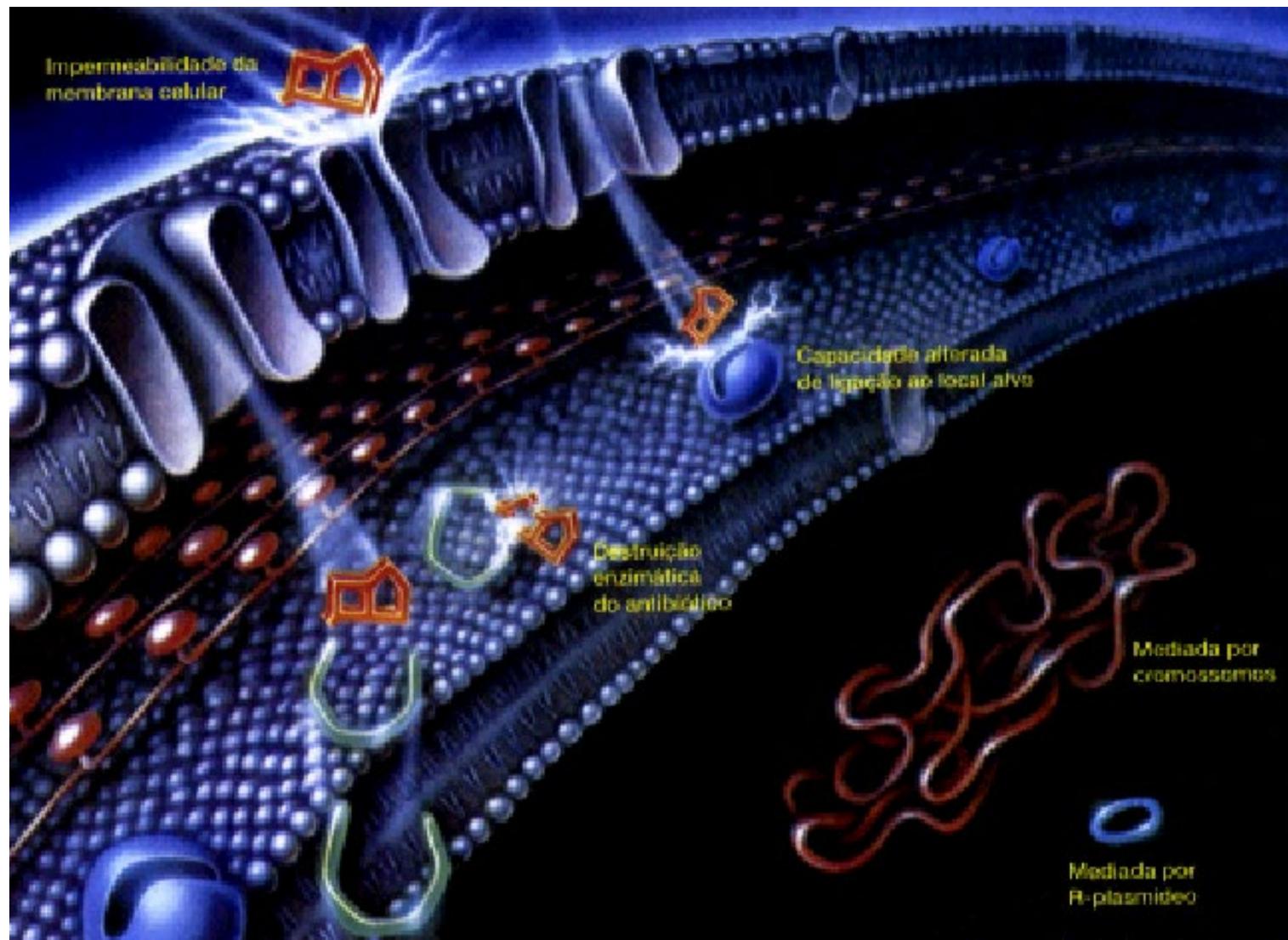
Priority 2: HIGH

- *Enterococcus faecium*, vancomycin-resistant
- *Staphylococcus aureus*, methicillin-resistant, vancomycin-intermediate and resistant
- *Helicobacter pylori*, clarithromycin-resistant
- *Campylobacter* spp., fluoroquinolone-resistant
- *Salmonellae*, fluoroquinolone-resistant
- *Neisseria gonorrhoeae*, cephalosporin-resistant, fluoroquinolone-resistant

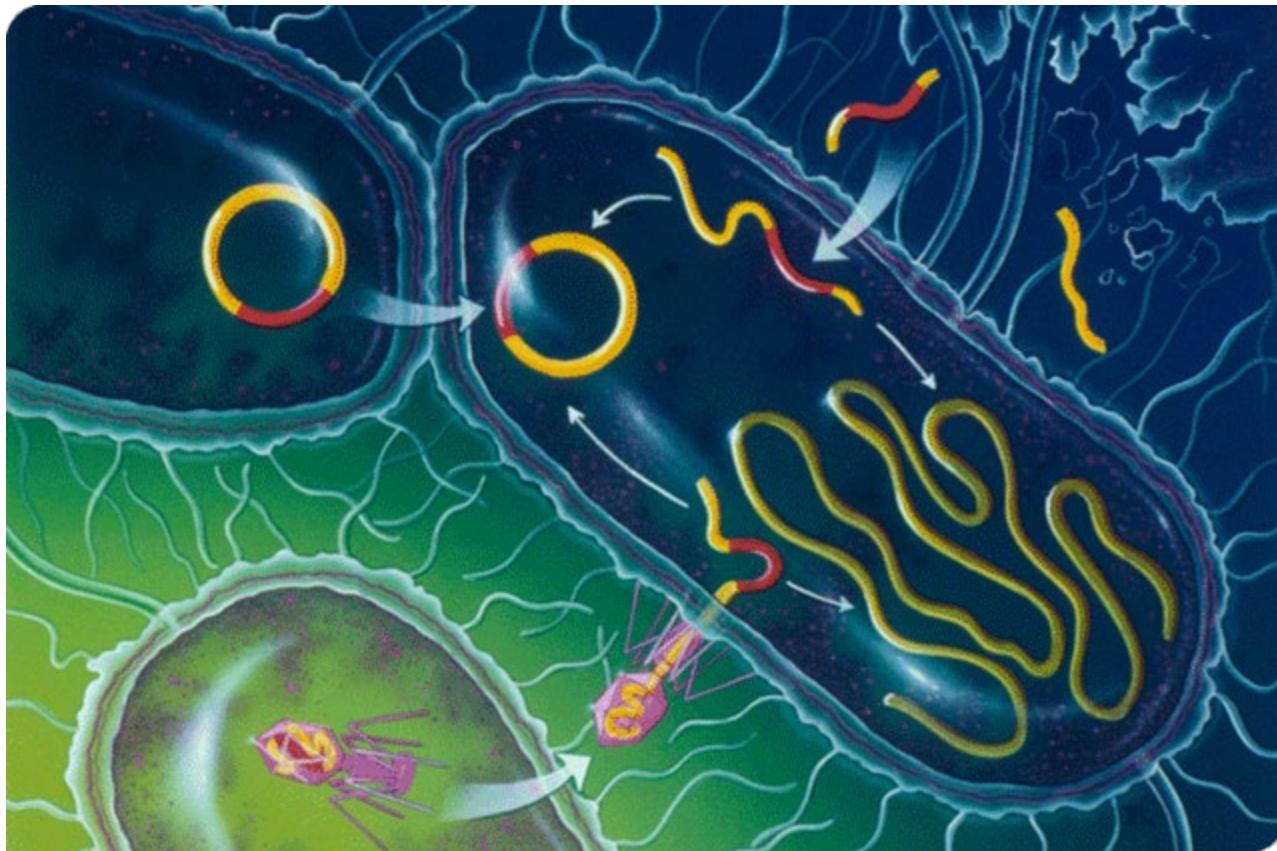
Priority 3: MEDIUM

- *Streptococcus pneumoniae*, penicillin-non-susceptible
- *Haemophilus influenzae*, ampicillin-resistant
- *Shigella* spp., fluoroquinolone-resistant

Resistencia a los beta-lactámicos (BLEE/Carba)

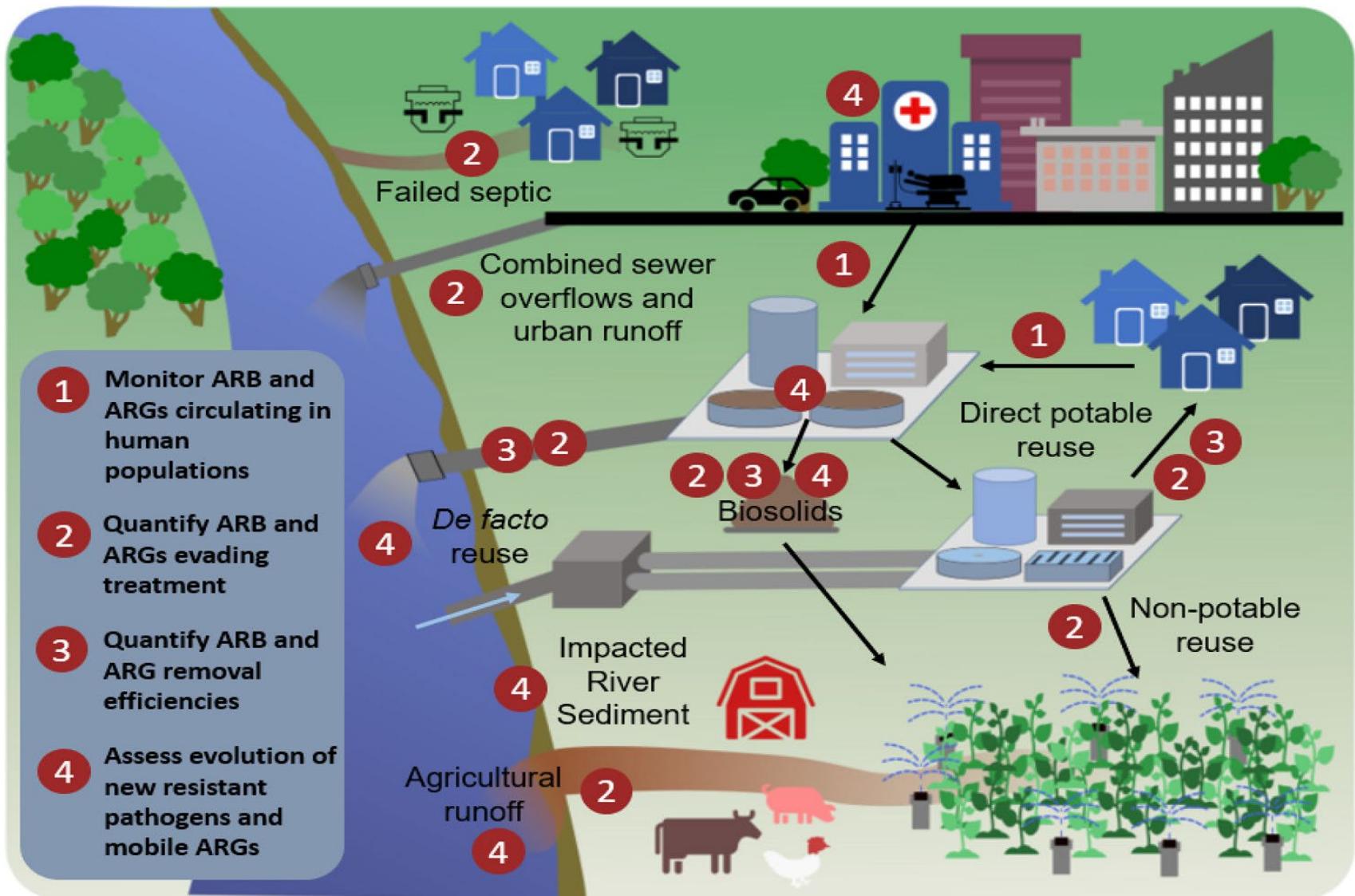


Mobilización genética



1. **Conjugación**
2. **Transformación**
3. **Transducción**

Vigilancia de la RAM



1. Vigilancia: bacterias bioindicadoras

Available at www.veterinaryworld.org/Vol.15/April-2022/33.pdf

Table-1: Bacterial species for inclusion in AMR surveillance programs in animals.

Bacterial Species	Category	Associated Animal Species	Reference
Methicillin-resistant <i>Staphylococcus pseudintermedius</i>	Animal-only pathogens	Companion animals	[25]
<i>Mannheimia haemolytica</i> , <i>Pasteurella multocida</i> , and <i>Histophilus somni</i>	Animal-only pathogens	Cattle	[24]
<i>Actinobacillus pleuropneumoniae</i> , <i>Haemophilus parasuis</i>	Animal-only pathogens	Pig	[26]
<i>Pasteurella multocida</i>	Animal-only pathogens	Cattle, pig	[21]
Enterotoxigenic <i>E. coli</i>	Animal-only pathogens	Pig, calves	[21]
<i>Salmonella</i> spp.	Animal-only pathogens	Slaughtered food animals	[21]
Methicillin-resistant <i>Staphylococcus aureus</i> and extraintestinal pathogenic <i>E. coli</i>	Zooanthroponotic pathogens	Dog, cat, horse	[21]
<i>Salmonella</i> spp. and <i>Campylobacter</i> spp.	Zoonotic foodborne pathogens	Cattle, pig and poultry	[21]
<i>Enterococcus</i> spp. and <i>E. coli</i>	Indicator bacteria	Healthy livestock and poultry	[27]

Renibacterium salmoninarum

Streptococcus iniae

Flavobacterium psychrophilum

Aeromonas spp.

Vibrio spp.

Edwarsiella

Flavobacterium spp.

Piscirickettsia salmonis

¿Acuicultura?

Vet World. 2022 Apr;15(4):1066-1079.

2. Vigilancia: cultivo de bacterias

Available at www.veterinaryworld.org/Vol.15/April-2022/33.pdf

Table-2: Media used for culture of bacteria recommended for AMR surveillance programs in animals.

Bacterial Species	Pre-enrichment		Selective enrichment		Isolation		Reference
	Media	Incubation	Media	Incubation	Media	Incubation	
<i>Staphylococcus</i> spp.	-	-	-	-	Baird-Parker agar	35-37°C (45-48 h)	[38]
<i>Mannheimia haemolytica</i>	-	-	-	-	Blood agar	37°C (24 h)	[29]
<i>Pasteurella multocida</i>	-	-	-	-	Blood agar	37°C (24 h)	[29]
<i>Escherichia coli</i>	Lactose broth	35±2°C (24 h)	EC broth	44.5°C (24 h)	L-EMB, EMB agar	35±2°C (24 h)	[39]
<i>Salmonella</i> spp.	Lactose broth	35°C (24 h)	TT broth	35±2.0°C (24 h)	XLD agar	35°C (24 h)	[40]
	BPW	35±2°C (24 h)	RV broth	42°C (24 h)	BS agar HE agar	35°C (24 h) 35°C (24 h)	
<i>Campylobacter</i> spp.	Bolton broth	37°C (4 h)	Bolton broth	42°C (48 h)	mCCDA AHB agar	37-42°C (24-48 h) 37-42°C (24-48 h)	[41]

BPW=Buffered peptone water, TT=Tetrathionate, RV=Rappaport-Vassiliadis, L-EMB=Levine's eosin-methylene blue, EMB=Eosin-methylene blue, XLD=Xylose lysine desoxycholate, BS=Bismuth sulfite, HE=Hektoen enteric, mCCDA=Modified campy blood-free agar, AHB=Abeyta-Hunt-Bark

3. Vigilancia: antibiótico (biomarcador)

Table-3: Suggested antimicrobials for inclusion in AMR surveillance programs in animals.

Antibiotic class	Antibiotic	Target bacterial species	Reference
Aminoglycosides	Gentamicin	<i>Salmonella</i> , <i>E. coli</i> , <i>Campylobacter</i> , <i>Enterococcus</i> , <i>Staphylococcus</i>	[11,21,28,29]
Amphenicols	Streptomycin Chloramphenicol	<i>Campylobacter</i> , <i>Enterococcus</i> <i>Salmonella</i> , <i>E. coli</i> , <i>Enterococcus</i> , <i>Staphylococcus</i>	
Second generation cephalosporins	Cefoxitin	<i>Salmonella</i> , <i>E. coli</i> , <i>Staphylococcus</i>	
Third generation cephalosporins	Cefatoxime Ceftriaxone Ceftazidime	<i>Salmonella</i> , <i>E. coli</i> <i>Salmonella</i> , <i>E. coli</i> <i>Salmonella</i> , <i>E. coli</i>	
Quinolones	Ciprofloxacin	<i>Salmonella</i> , <i>E. coli</i> , <i>Campylobacter</i> , <i>Enterococcus</i> , <i>Staphylococcus</i>	
	Nalidixic acid	<i>Salmonella</i> , <i>E. coli</i> , <i>Campylobacter</i>	
Rifamycins	Pefloxacin	<i>Salmonella</i> , <i>E. coli</i>	
Sulfonamides	Rifampicin	<i>Staphylococcus</i>	
	Sulfisoxazole	<i>Salmonella</i> , <i>E. coli</i> , <i>Staphylococcus</i>	
	Trimethoprim-sulfamethoxazole	<i>Salmonella</i> , <i>E. coli</i> , <i>Staphylococcus</i>	
Trimethoprim	Trimethoprim	<i>Salmonella</i> , <i>E. coli</i> , <i>Staphylococcus</i>	
Tetracyclines	Tetracycline	<i>Salmonella</i> , <i>E. coli</i> , <i>Campylobacter</i> , <i>Enterococcus</i> , <i>Staphylococcus</i>	
	Doxycycline	<i>Campylobacter</i>	
Carbapenems	Imipenem	<i>Salmonella</i> , <i>E. coli</i>	
	Meropenem	<i>Salmonella</i> , <i>E. coli</i>	
Polymyxins	Colistin	<i>Salmonella</i> , <i>E. coli</i>	

4. Vigilancia: método fenotípico

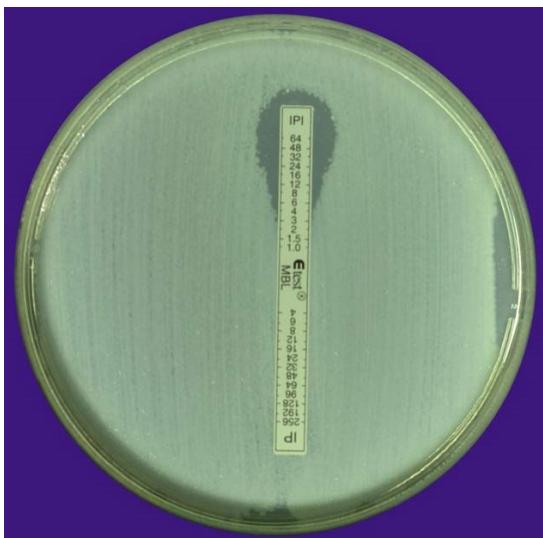
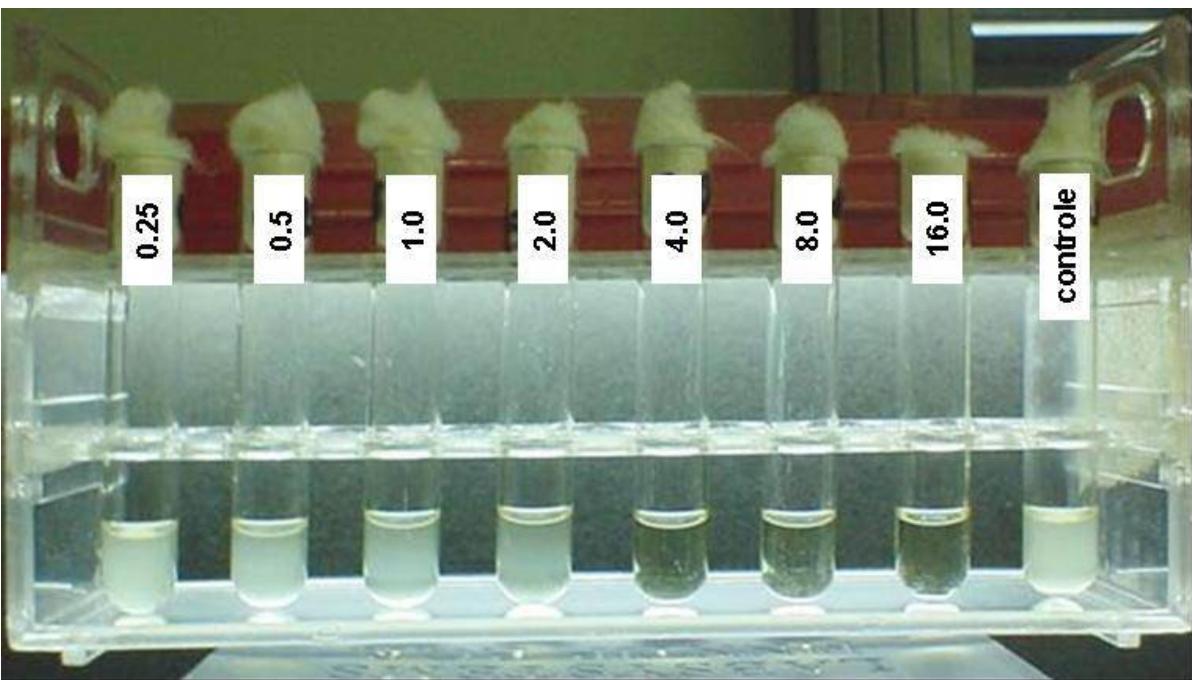
Table-4: Phenotypic antimicrobial susceptibility test methods commonly used in laboratories.

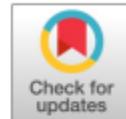
Name of the AST	Nature of the AST	Media used	Time required (h)	Antibiotics that can be tested	Reference
Disk diffusion method	Phenotypic, qualitative	MHA	18-24	All antibiotics except colistin	[12,46]
Broth dilution method			24	All antibiotics	[12,45,46]
Agar dilution method			24	All antibiotics except colistin and sulfa drugs	[45]
Etest®			24	All antibiotics	[46]
Sensititre™			18-24	All antibiotics	[46]
Vitek 2®			4-10	All antibiotics	[46]
BD Phoenix			6-16	All antibiotics	[46]
MicroScan®			4.5-7	All antibiotics	[46]

MHA=Mue

=Aspartate aminotransferase

4. Vigilancia: método fenotípico





Optimizing a Screening Protocol for Potential Extended-Spectrum β -Lactamase *Escherichia coli* on MacConkey Agar for Use in a Global Surveillance Program

Megan E. Jacob,^a Shivaramu Keelara,^a Awa Aidara-Kane,^b Jorge R. Matheu Alvarez,^b Paula J. Fedorka-Cray^a

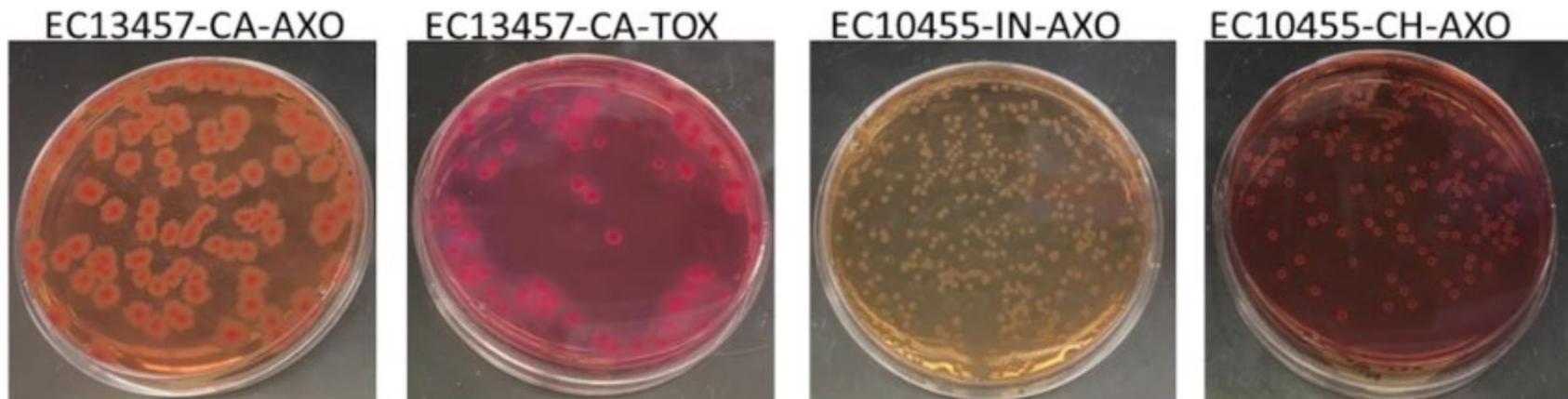
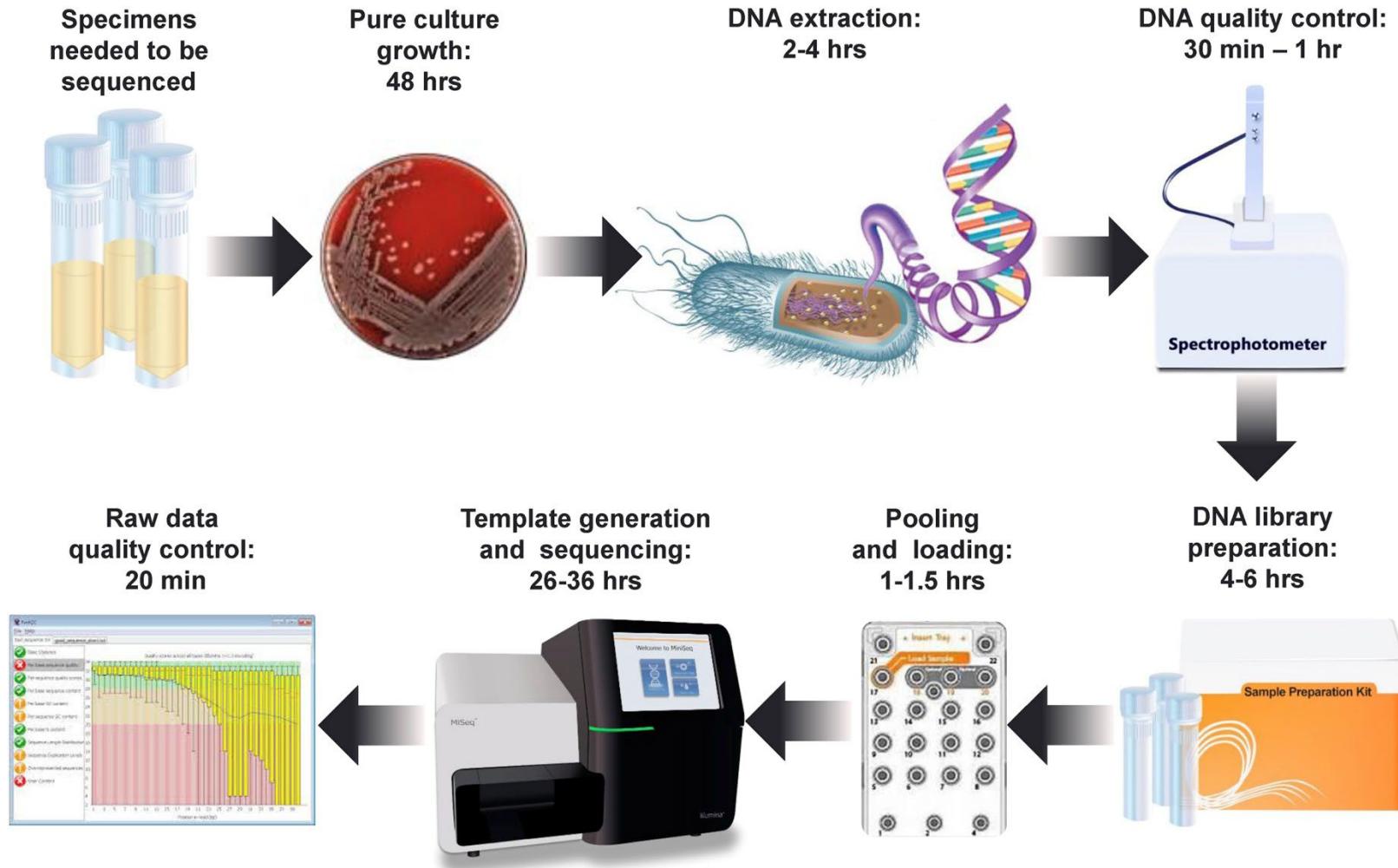
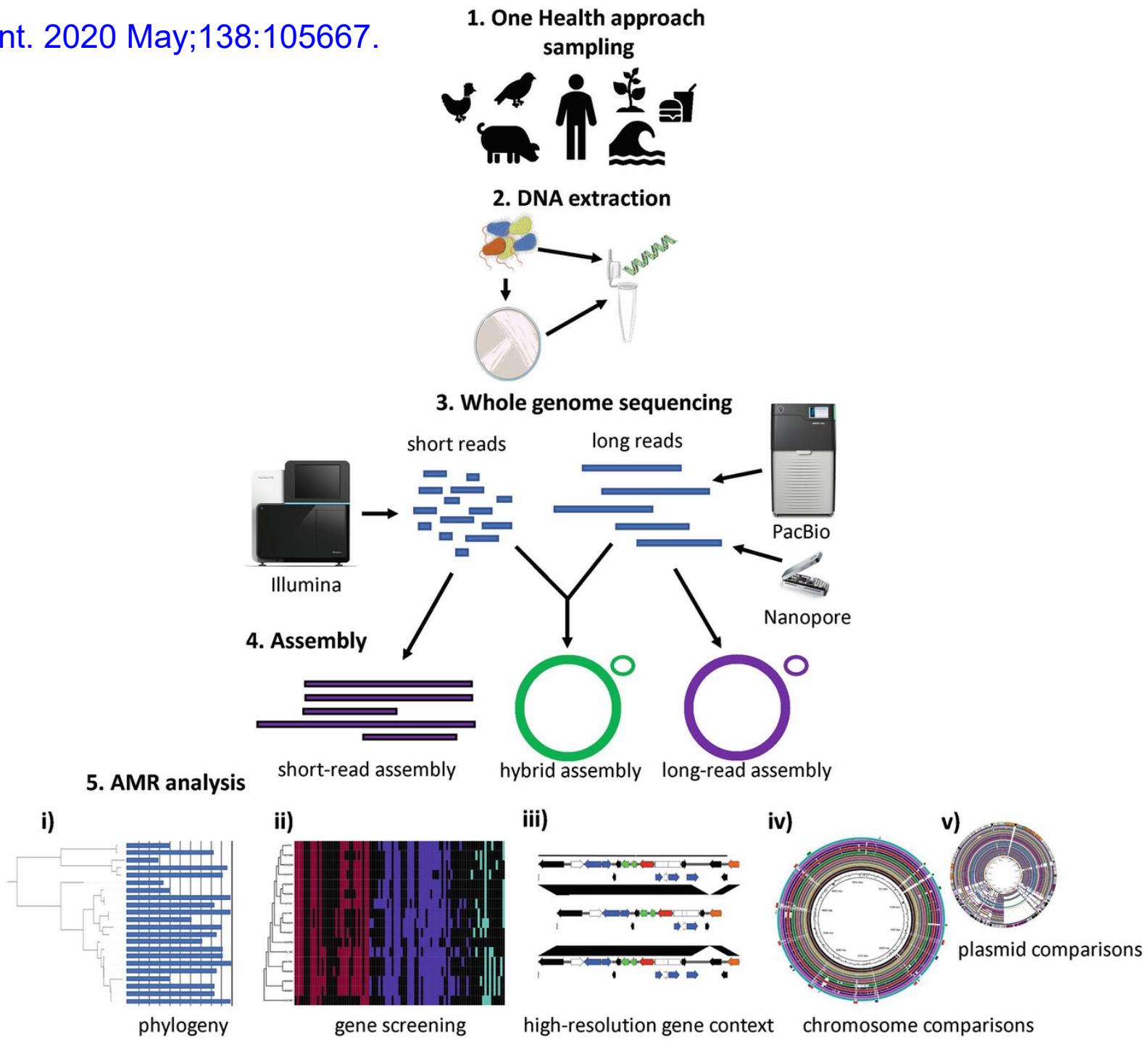


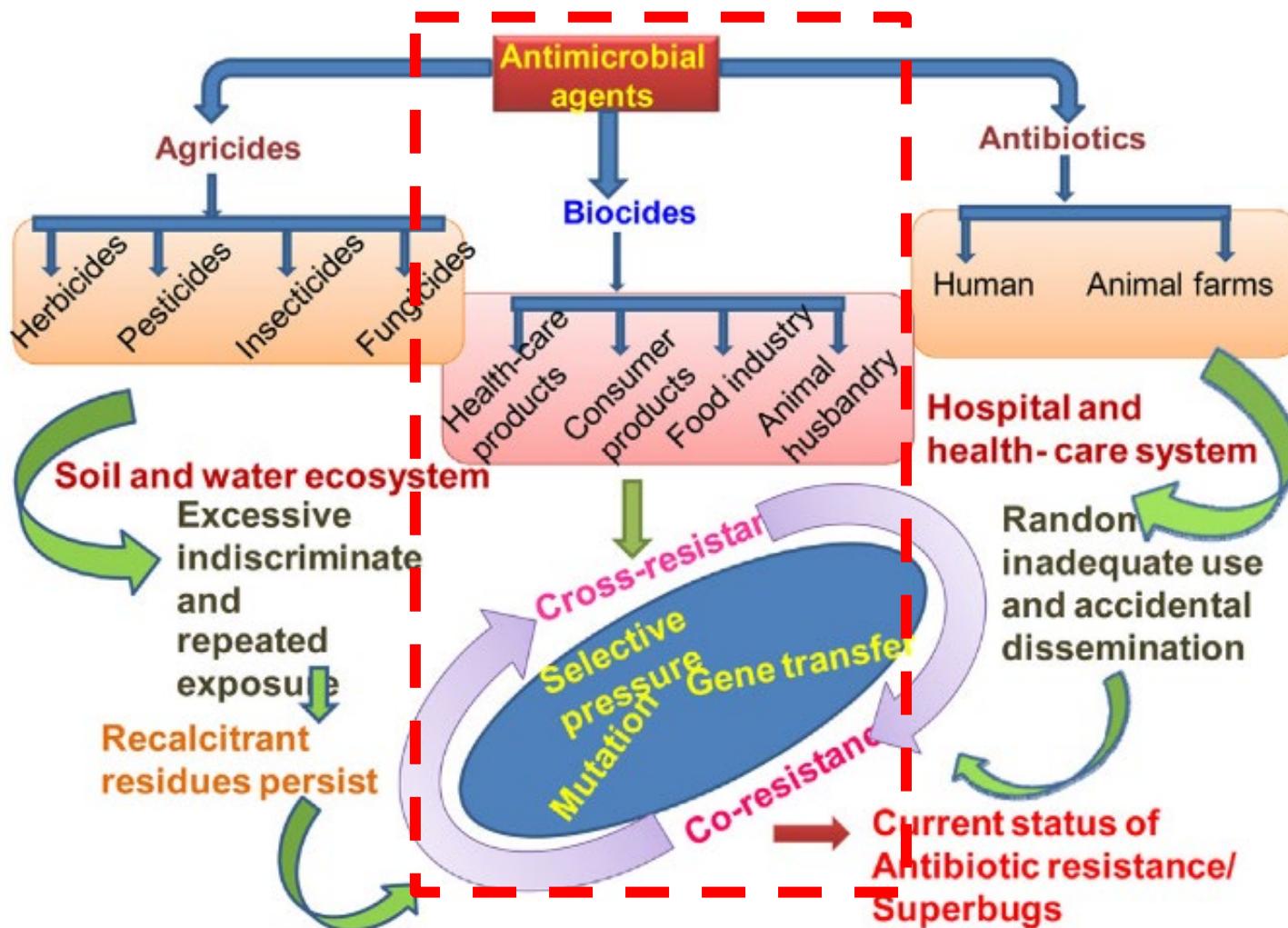
FIG 2 Phenotypic appearance of pure cultures of *Escherichia coli* (EC) 13457 and *E. coli* 10455 on MacConkey agar manufactured in Canada (CA), India (IN), and China (CH) supplemented with 4 μ g/ml either cefotaxime (TOX) or ceftriaxone (AXO).

5. Vigilancia genómica/epidemiológica





Desinfectantes y Biocidas: resistencia/tolerancia y resistencia cruzada/co-resistencia ATM



Paul et al., 2019. *Ecotoxicol Environ Saf.* 174:601-610.
Ansari et al., JAC *Antimicrob Resist.* 2021;3:dlab038

Gracias por la atención!



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